

Non Technical Description

Cancer of the ovary is responsible for more than 12,000 deaths annually in the States. The disease is usually only diagnosed at a stage when the tumor has spread to involve structures in the abdominal cavity. The standard treatment for ovarian cancer involves surgical removal of the larger tumor masses combined with a combination of chemotherapy drug. Although new chemotherapy drugs such as taxol are effective, complete responses are few, and toxicity is high. It therefore remains necessary to develop new treatment strategies to improve the survival and quality of life of patients who are afflicted with ovarian cancer.

The results of studies conducted in our laboratories and others have shown that a particular cell type called the tumor infiltrating lymphocyte (TIL) can be isolated from the cancerous tumors that involve the lining of the abdominal cavity and the surface of the intestine. When these cells are removed from the body and are grown together with the patient's tumor cells in tissue culture, along with a drug called recombinant interleukin-2 (rIL-2), the TILs will frequently increase to larger numbers and develop an increased capacity to kill tumor cells obtained from the patient. rIL-2 is a protein substance that is almost identical to a natural substance that is found in the body and is produced in very small amounts by the cells of the body's immune system called lymphocytes. We are currently growing large numbers of TILs for the experimental treatment of patients with advanced ovarian cancer in a clinical trial. This type of treatment has been called adoptive cellular immunotherapy. In this trial we purify a particular type of TIL called "CD8+ TIL" which appear to be responsible for the killing of tumor cells in tissue culture in certain cases.

This gene marker protocol will address an important scientific question: whether ovarian TIL-derived T cells that are expanded ex vivo in rIL-2 for intraperitoneal injection, concentrate within metastatic tumors of the abdominal cavity. This information will be helpful in the development of improved therapies for ovarian cancer. In the past this type of question could only be addressed by placing a radioactive label onto the cells and trying to trace the cells using a gamma camera held over the patient. These techniques remain in use but are not sensitive enough to address the questions that are being posed.

Recently, use has been made of defective mouse viruses that have been genetically altered in the laboratory so that they are rendered incapable of multiplying in the patient. These viruses have been used to transfer new genetic information into cells. The gene transfer approach is being used to introduce a stable marker into cells that are being returned to the body for marker studies or in the development of new therapy approaches. Appropriate concerns have been expressed regarding utilization of these defective viruses in patients. Marked improvements in design and in quality control are contributing nevertheless, to safer and more efficient clinical applications.

In our study we intend to use the defective virus GINA, to transfer the gene for marker DNA. This marker gene is not present in human or mammalian cells. The gene encodes for an enzyme called neomycin phosphate transferase (NPT). The defective virus and related forms have been used on a number of occasions to treat patients without reports of toxicity.

Patients who are enrolled in a study to be treated with TIL will be eligible for the marker protocol. During the growth of TIL in culture a bacterial "marker" gene will be inserted into a portion of cultured TIL cells using a defective mouse retrovirus so that they can be traced after reinjection into the patient. Following injections of a mixture of gene marked and unmarked cells, samples of blood (about 10 cc each) and peritoneal fluid (approximately 100 cc) will be obtained on days 4, 11, 18 and at one month after receiving the TIL injection. One month after receiving the TIL, patients will undergo a laparoscopy to evaluate their responses to the treatment. At this time biopsies will be obtained from tissues suspected of harboring cancer. Biopsies will also be obtained from certain normal tissues, e.g., peritoneum (lining of abdominal cavity) and lymph node (if easily accessible). These biopsies will be obtained to determine if the TIL have migrated into the tumor. This information might be useful for the development of future treatment strategies in other patients with ovarian cancer. The laparoscopic surgery will be performed to evaluate the response to the treatment and not for the marker study alone. All patients may receive the retrovirus modified cells.